

UK Standards for Microbiology Investigations

Laboratory diagnosis of syphilis



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Acknowledgments

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Amendment table

Each UK SMI document has an individual record of amendments. The amendments are listed on this page. The amendment history is available from <u>standards@ukhsa.gov.uk</u>.

Any alterations to this document should be controlled in accordance with the local document control process.

Amendment number/date	7/23.05.25	
Issue number discarded	3	
Insert issue number	3.1	
Section(s) involved	Amendment	
	This is an administrative point change.	
	The content of this UK SMI document has not changed.	
	The last scientific and clinical review was conducted on 19.12.2023.	
	Hyperlinks throughout document updated to Royal College of Pathologists website.	
Whole document	Public Health England replaced with UK Health Security Agency throughout the document, including the updated Royal Coat of Arms	
	Partner organisation logos updated.	
	Broken links to devolved administrations replaced.	
	References to NICE accreditation removed.	
	Scope and Purpose replaced with General and Scientific information to align with current UK SMI template.	
Section 10: Public health responsibilities of diagnostic laboratories	This section has been added to UK SMI templates to highlight the public health responsibilities that diagnostic laboratories have as part of their duties.	

Amendment number/date	6/18.12.23	
Issue number discarded	2.1	
Insert issue number	3.0	
Anticipated next review date*	18.12.26	
Section(s) involved	Amendment	
Title	The title has been changed from syphilis serology to Laboratory diagnosis of syphilis.	
Introduction	Included primary, secondary, latent and tertiary stages. Confirmatory treponemal test TPPA has been withdrawn from the UK in 2022 due to regulatory requirements. The diagnostic algorithm has been updated to remove TPPA and gives the option of using either TPHA/TPLA or a second EIA/CLIA as the confirmatory test.	
Algorithm/Tables	Treponemal serology screening and confirmation algorithm updated to remove TPPA and gives the option of using either TPHA/TPLA or a second EIA/CLIA as the confirmatory test. All interpreting and reporting tables have been restructured with all the common scenarios.	
References	References reviewed and updated.	

*Reviews can be extended up to 5 years where appropriate

1 General information

View general information related to UK SMIs.

2 Scientific information

View scientific information related to UK SMIs.

3 Scope of document

This UK Standards for Microbiology Investigation (UK SMI) document describes laboratory testing for diagnosis of *Treponema pallidum* infection. It is concerned with diagnosis of syphilis including primary, secondary, latent and tertiary syphilis including central nervous system (CNS) and congenital infections.

Refer to <u>UK SMI S 6: Infectious syndromes affecting the genitourinary tract and reproductive</u> organs for further information regarding clinical presentations of sexually transmitted infections, and associated tests.

UK SMIs should be used in conjunction with other relevant UK SMIs.

4 **Definitions**

- TPPA Treponema pallidum particle agglutination assay
- TPHA Treponema pallidum haemagglutination assay
- TPLA Treponema pallidum latex agglutination assay
- EIA Enzyme immunoassay
- CLIA Chemiluminescent immunoassay
- RPR Rapid plasma reagin
- VDRL Venereal disease research laboratory

5 Introduction

Syphilis is a sexually transmitted disease caused by the bacterium *Treponema pallidum* subsp. *pallidum* (1).

Syphilis is typically transmitted by direct contact with an infectious lesion through genital or extra genital sites (anal, rectal and oral). Transmission can occur in utero at any stage of pregnancy, although the risk is increased in the second half of pregnancy, and at the time of birth through contact with maternal lesions in the birth canal (1).

Rare routes of transmission include injecting drugs and blood transfusion (1).

Syphilis is grouped into primary, secondary, latent or tertiary stage. Neurosyphilis can occur at any stage of infection (2).

- primary stage- ulcer (usually painless) or chancre found at the inoculation site usually located on the genitals, rectum, tongue, or lips, which occurs 10-90 days after exposure (1,2)
- secondary stage- signs and symptoms include a skin rash marked by red or reddishbrown macules on the palms and soles or other parts of the body, mucocutaneous lesions, lymphadenopathy, anorexia, fever, sore throat, headaches, weight loss, fatigue, mucous patches, condylomata lata and alopecia. This occurs 2-10 weeks after the chancre appears
- latent stage- No signs or symptoms are present. Early latent is within 1- 2 years of infection, and late latent thereafter. In some patients, latent syphilis ends with the development of tertiary disease
- tertiary stage- signs include gummata, cardiac or neurological manifestations. This stage generally occurs 10-20 years after infection

Currently primary, secondary and early latent syphilis is increasing among gay, bisexual or other men who have sex with men (GBMSM), and heterosexual people (3). In 2022, the number of diagnosed cases reported were 8,692. This increased by 15.2% compared to 2021 where 7,543 diagnosed cases were reported. This is the largest annual number of syphilis diagnoses since 1948 (4).

Syphilis shares many clinical features with other treponemal and non-treponemal diseases. *T. pallidum* subsp. *pertenue* (yaws), and *T. pallidum* subsp. *endemicum* (bejel) are morphologically identical subspecies of *T. pallidum* (5). Therefore, laboratory test results must be considered together with the clinical and geographical background of the patient because the serological assays used for syphilis testing also detect antibody raised in response to endemic treponematoses (5,6). As a precaution an individual with positive treponemal serology should be investigated and treated for syphilis unless previous treatment can be documented (7).

In suspected early primary syphilis, a sample should ideally be taken from the lesion for treponemal PCR (8). Examination for treponemes by dark ground microscopy may be undertaken although PCR is preferable when investigating lesions likely to be contaminated with commensal treponemes such as oral lesions (7,9,10).

Most UK laboratories used a *T. pallidum* EIA/CLIA screening test (test 1) followed by TPPA as the confirmatory treponemal test until this was withdrawn from the UK in 2022 due to regulatory requirements. The diagnostic screening algorithm in this UK SMI has been updated to remove TPPA and gives the option of using either TPHA/TPLA or a second EIA/CLIA as the confirmatory test (test 2).

6 Treponemal serology screening and confirmation algorithm

Refer to section 8 for the neonatal algorithm.

6.1 Footnotes

- a. At least one test should be performed using the primary tube.
- b. False negative screening results may be seen in immunocompromised individuals. Negative results within 3 months of infection cannot exclude early syphilis.
- c. Treponemal IgM tests lack sensitivity and specificity and should not be used to stage disease, diagnose reinfections or determine the duration of treatment. A positive IgM result may be useful if primary syphilis is suspected. Results can only be interpreted in association with other treponemal and non-treponemal antibody test results and clinical information. True positive results may reflect recent or active infection but note that IgM reactivity can persist for 12 18 months even after adequate treatment of infection (7).
- d. A second treponemal test is used to confirm screen positive results and identify false positives. Most CLIA/EIAs use one or more recombinant treponemal antigens. They are sensitive but may have poor specificity. Where possible, laboratories should use a second treponemal assay that uses different antigen targets to the screening assay to

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inform the likelihood of false positive results. Conversely, TPHA/TPLA may be less sensitive than some CLIA/EIAs resulting in true screen positive results which fail to confirm. It is important to request a second sample 2 weeks later if early syphilis is suspected.

- e. Prozone effect (high antibody titres) leading to false negative results may be observed in secondary syphilis or early latent syphilis.
- f. RPR should also be repeated on the day of commencing treatment so that the highest titre is documented.

6.2 Interpreting and reporting laboratory results for treponemal serology and NAAT testing

Note that the table of comments is a guide, and that clinical details and previous serological results should always be considered when interpreting treponemal serology results.

The table cannot cover all serological profiles but should cover most of those encountered in clinical practice. A full repertoire of tests for final interpretation may include referral tests, depending on the local laboratory test repertoire.

Note that the quotation marks indicate the report comment.

Treponemal test 1	Treponemal test 2	RPR	Report comment	Notes
(EIA/CLIA)	(EIA/CLIA/ TPHA/TPLA)			
Positive	Positive	≤1:16 or including negative	"Consistent with treponemal infection at some time. Active infection is not excluded. Please send a further sample if this is a new diagnosis. Serology results should be interpreted according to clinical presentation." Refer to notes section.	This would be consistent with a recent infection if seroconversion, or a four- fold rise in RPR titre was seen in comparison to an earlier sample, or if there were clinical signs suggesting early syphilis.
			If first sample: "Result may be due to non- specific cross-reactivity or early infection." Please send a further sample in two weeks to exclude early syphilis. Serology results should	EIA/CLIAs (and RPR is negative) consider reviewing level of reactivity in treponemal tests.
			be interpreted according to clinical presentation and history of risk. If same profile on repeat sample (at least two	Laboratories need to establish what constitutes a low level reactive result with each test in use, according to local data.
			weeks later): "Persistent reactivity in treponemal tests may be non-specific. Serology results should be interpreted according to clinical presentation and history of risk."	Low level reactivity in both treponemal assays may be consistent with treponemal infection but could possibly be due to non-specific cross-reactivity. Interpret in the context of clinical
				presentation and history of risk. If the same profile is seen on repeat testing and the patient has clinical features or risk factors for syphilis, consider treponemal IgG immunoblot testing.

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Treponemal test 1 (EIA/CLIA)	Treponemal test 2 (EIA/CLIA/ TPHA/TPLA)	RPR	Report comment	Notes
Positive	Positive	Positive at >1:16	"Consistent with recent or active treponemal infection. Please send a further sample if this is a new diagnosis. Serology results should be interpreted according to clinical presentation."	An RPR of >1:16 is suggestive of active or recent infection, or re- infection. If this is a follow-up sample, review previous results and report changes in RPR titre. Follow-up RPR testing should be according to BASHH guidelines (1).
Negative	Negative	Positive (any titre)	"Isolated RPR reactivity is likely to reflect non- specific reactivity. Please send a repeat sample in two weeks to exclude recent infection. Serology results should be interpreted according to clinical presentation."	None.
Negative	Negative	Negative (if done)	"No serological evidence of treponemal infection. In suspected primary syphilis, consider testing a further sample taken at least two weeks after onset of symptoms to account for the possible seronegative window in early cases. In cases of recent contact, retest after 3 months or earlier if compatible symptoms develop."	Antibody responses may be reduced in the immunosuppressed.

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Treponemal test 1	Treponemal test 2	RPR	Report comment	Notes
(EIA/CLIA)	(EIA/CLIA/ TPHA/TPLA)			
Positive or equivocal	Negative	Negative	If first sample: "Result may be due to non- specific cross-reactivity or early infection. Please send a further sample in two weeks to	Evaluate level of reactivity in treponemal test. Laboratories need to establish what constitutes a low level
Negative	Positive or equivocal	Negative	exclude early syphilis. Serology results should be interpreted according to clinical presentation and history of risk." If same profile on repeat sample (at least two weeks later): "Persistent reactivity in one treponemal test is probably non-specific. Serology results should be interpreted according to clinical presentation and history of risk."	reactive result with each test in use, according to local data. If the same profile is seen on repeat testing and the patient has clinical features or risk factors for syphilis, or high level of reactivity, consider performing or referral for treponemal IgG immunoblot testing.
Positive or equivocal	Negative	Positive (any titre)	If first sample: "Result may be due to non- specific cross-reactivity or early infection. Please send a further sample in two weeks to	This is an unusual profile. Evaluate level of reactivity in treponemal test and RPR titre.
Negative	Positive or equivocal	Positive (any titre)	exclude early syphilis. Serology results should be interpreted according to clinical presentation and history of risk."	If RPR titre is high, consider treating. If the same profile is seen on repeat testing, perform or refer for treponemal IgG immunoblot testing for resolution of antibody status.

NAAT Report comment		
Detected	T. pallidum detected consistent with active syphilis infection	
Not detected	T. pallidum not detected. Review syphilis serology in light of clinical presentation	

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7 Diagnosis of neurosyphilis

T. pallidum commonly invades the central nervous system at an early stage of infection and may or may not produce symptoms. The diagnosis is based on clinical findings with positive serological tests.

Symptomatic infection can present early as aseptic meningitis or later as meningovascular syphilis or parenchymal late neurosyphilis including general paresis and tabes dorsalis (11). In the preantibiotic era some 20% of infected individuals developed symptomatic neurosyphilis. Early treatment with penicillin markedly reduces the risk of progression from asymptomatic to symptomatic CNS infection.

No single test can diagnose neurosyphilis and similarly no CSF result can definitively exclude a diagnosis of neurosyphilis. Local validation must be performed for treponemal and non-treponemal serology tests performed on CSF. Diagnosis of neurosyphilis requires consideration of the history (including risk factors, treatment history and HIV status), clinical findings, and CSF microscopy and protein level, together with blood and CSF treponemal serology results. CSF protein is variably raised in neurosyphilis depending on the stage of infection. CSF pleocytosis, when present, is lymphocytic. An average of 25-75 cells X 10^6/L is found in tabes dorsalis and general paresis. However, the CSF is acellular in 10% of cases of tabes dorsalis.

Consider the following for CSF samples:

- if the peripheral blood is negative for treponemal antibodies there is no need to test a CSF sample
- testing of CSF should be considered in patients with treponemal infection and neurological signs or symptoms (1). Blood contamination of CSF should be minimised. A matched serum sample should be taken to compare antibody levels with CSF levels

7.1 Treponemal serology in neurosyphilis

Much of the original work on serological diagnosis of syphilis was performed using VDRL as the non-treponemal test for CSF. However, changes in practice now mean that RPR is more commonly used. Following the withdrawal of the TPPA assay from the UK, TPHA maybe performed alongside RPR. If CSF RPR is negative, consider performing TPHA if available.

Consider the following for neurosyphilis:

- CSF RPR is an insensitive test for neurosyphilis being positive in only about 50% of cases (1,12). A positive RPR, in the absence of evidence of blood contamination of the CSF sample, is diagnostic of neurosyphilis (1)
- a negative CSF TPHA makes a diagnosis of neurosyphilis unlikely. A positive CSF TPHA test is highly sensitive for neurosyphilis but lacks specificity because reactivity may be caused by transudation of immunoglobulins from the serum into the CSF. CSF TPHA titres can help to distinguish between higher antibody levels associated with neurosyphilis due to intrathecal antibody production and lower levels due to passive transfer from the blood. A CSF TPHA titre >1:320 is sensitive

and specific for neurosyphilis and may be helpful in supporting the diagnosis of neurosyphilis when the CSF RPR is negative

- non-treponemal test results on peripheral blood can help to predict, or exclude, neurosyphilis: a negative RPR virtually excludes neurosyphilis, whereas RPR ≥1:32 increases the likelihood of neurosyphilis (approximately 11-fold in patients without concurrent HIV infection and 6- fold in the HIV-infected individual) (13,14)
- supporting evidence for the use of *T. pallidum* PCR for diagnosing neurosyphilis is weak; studies are generally small and heterogenous due to lack of a diagnostic gold standard. In studies using a positive CSF VDRL to diagnose neurosyphilis, the sensitivity of the PCR varied between 40% and 70% and specificity between 60% and 100% (15)

Following treatment for neurosyphilis, any CSF pleocytosis should have decreased within six months and CSF should be normal within two years (except for persistent positive treponemal specific antibody tests) (1).

8 Early congenital syphilis testing algorithm

The diagnosis of congenital syphilis can be very difficult; most infected neonates appear normal at birth and passive transfer of maternal syphilis antibodies may cause reactive neonatal serology in the uninfected infant.

A recent review of congenital syphilis cases in England found that most had been born to mothers who had negative antenatal screening results, meaning they became infected with syphilis later during their pregnancy and were undiagnosed and untreated before giving birth. The majority of mothers were diagnosed following their symptomatic infant's diagnosis (16).

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8.1 Footnotes relating to early congenital syphilis

a. Symptomatic baby with signs suggesting possibility of congenital syphilis. Early congenital syphilis manifests within two years of birth. Symptoms might include: snuffles, perioral fissures, hepatomegaly and jaundice, cataracts, growth retardation, rash, mucous patches and condylomata lata.

It is advisable to take and compare a contemporaneous blood from the mother when investigating the baby's serum.

- b. All children born to mothers with confirmed positive treponemal serology require clinical evaluation and syphilis serology tests, with the following exceptions:
 - maternal biological false-positive serology
 - maternal syphilis cured prior to this pregnancy

Women with positive syphilis serology who have been adequately treated prior to this pregnancy may have ongoing risk factors for infection. Review of cases of vertical transmission of syphilis have shown that some women acquire syphilis and develop primary infection after an initial negative screening result. This highlights the importance of offering repeat testing to women at risk of exposure to syphilis and/or who have relevant symptoms at any stage of pregnancy.

Passively transferred maternal non-treponemal antibodies should decline by three and be negative by six months of age. Passively transferred treponemal antibodies should decline by 18 months of age.

Infants should be tested at birth and at three months of age, and then the RPR repeated at three monthly intervals until negative (1). If titres remain stable or increase evaluate and treat for congenital syphilis (1).

Note that when maternal syphilis is acquired late in the pregnancy antibodies might not be present in mother or baby at birth (17).

- c. Serological tests should be performed on baby's blood (not the cord blood). Treponemal IgM test should be the priority on small volume samples.
- d. Treponemal antibody test can be EIA, CLIA, TPLA or TPHA. There is no need to confirm with a 2nd treponemal test.
- e. Suitable samples for PCR include nasal discharge, naso-pharyngeal aspirate, throat swabs, lesion swabs, blood and CSF. If placental tissue is available this may also be tested by PCR. In the case of foetal loss where a post-mortem is performed, suitable samples for PCR (in addition to those above) include liver, lung and spleen tissue samples. NAAT testing maybe performed locally or can be discussed with a reference laboratory.
- f. Testing of siblings should be carried out when a maternal or a congenital syphilis diagnosis is made (1).

8.2 Interpreting and reporting laboratory results for early congenital syphilis

lgM	RPR	Treponemal test (EIA/CLIA/TPLA/TPHA)	Report comment	Notes
Positive	Positive (any titre)	Positive/ equivocal	"Consistent with congenital infection. Please repeat to confirm. Consider treponemal PCR on suitable samples."	Note that IgM false positives and false negatives may occur, so results must always be interpreted in conjunction with the maternal serology results and clinical history.
Positive	Negative	Negative	If mother has acquired syphilis late in pregnancy it is possible that the baby may be treponemal antibody negative around the time of birth: "Possible congenital syphilis. Please repeat to confirm and send samples	Note that IgM false positives and false negatives may occur, so results must always be interpreted in conjunction with the maternal serology results and clinical history.
			for treponemal PCR."	
			"No conclusive evidence of congenital syphilis. The IgM reactivity is likely to be false. Please test a further sample to confirm status. Verify maternal treponemal antibody. Consider treponemal PCR on suitable samples."	

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IgM	RPR	Treponemal test (EIA/CLIA/TPLA/TPHA)	Report comment	Notes
Negative	Negative	egative Negative	"No serological evidence of congenital syphilis." If baby <3 months old: "Repeat	If baby is >3 months old at time of testing, repeat sample is not necessary.
			sample at 3 months if mother acquired syphilis late in pregnancy."	
Negative	Positive with a titre ≥4	tive Positive/ equivocal a ≥4 s of ner	"Consistent with congenital syphilis. Please repeat to confirm. Consider treponemal PCR on suitable samples."	If no recent maternal RPR result is available for interpretation, add the comment:
	times that of mother			RPR result should be interpreted in comparison with maternal RPR titre. Please contact the laboratory to discuss.
Negative	Positive with a titre <4 times that of mother	Positive/ equivocal	"Serology profile suggestive of passively transferred maternal antibody. However, this must be interpreted in comparison with maternal serology and in a clinical context. Advise repeat RPR at 3 monthly intervals to monitor for changes in titres, or until RPR becomes negative."	Four-fold (or greater) difference in RPR titre has high sensitivity for the diagnosis of congenital syphilis. Note a lower RPR titre does not exclude the diagnosis; most infants with congenital syphilis have an RPR titre that is the same or one or two dilutions less than the maternal titre (18,19). If a mother acquires syphilis and seroconverts late in pregnancy the baby may be delivered prior to a mature antibody response. This results in a low RPR titre and negative IgM, even in the presence of congenital infection.

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lgM	RPR	Treponemal test (EIA/CLIA/TPLA/TPHA)	Report comment	Notes
Negative	Negative	Positive/ equivocal	If sample taken around the time of birth: "Serology profile suggestive of passively transferred maternal antibody. Advise repeat at 3 months to confirm negative RPR and exclude early congenital syphilis."	Note if RPR remains negative at 3 months of age it is unnecessary to repeat samples until treponemal tests are negative, which may persist until 12 months of age.
			Sample from 3 months of age: "Passively transferred maternal antibody. No further testing is necessary."	
Negative	Positive (any titre)	Negative	Repeat RPR on original sample and if repeat reactive report as "Probable false positive RPR. Repeat serology in 3 months"	

9 Safety considerations

The section covers specific safety considerations (20-39) related to this UK SMI, and should be read in conjunction with the general <u>safety considerations</u>.

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10 Public Health responsibilities of diagnostic laboratories

Diagnostic laboratories have public health responsibility as part of their duties. Amongst these are additional local testing, or referral, to further characterise the organism, as required, primarily for public health purposes e.g. routine cryptosporidium detection; serotyping or microbial subtyping; and a duty to refer appropriate specimens and isolates of public health importance to a reference laboratory.

Diagnostic laboratory outputs inform public health intervention, and surveillance data is required to develop policy and guidance, forming an essential component of healthcare. It is recognised that additional testing and referral of samples may entail some costs that has to be borne by the laboratory but in certain jurisdictions these costs are covered centrally.

Diagnostic laboratories should be mindful of the impact of laboratory investigations on public health and consider requests from the reference laboratories for specimen referral or enhanced information.

References

An explanation of the reference assessment used is available in the <u>scientific</u> <u>information section on the UK SMI website.</u>

- 1. M. Kingston PF, S. Higgins, O McQuillan and others UK National guidelines on the management of syphilis 2015. Internal Journal of STD & AIDs 2015.++
- Morshed M and others. British Columbia's Experience after Implementation of the Treponema pallidum Reverse Algorithm and PCR Detection, 2015 to 2020. Microbiology Spectrum 2022: volume 10, issue 3, pages e0068622.++ 10.1128/spectrum.00686-22
- 3. UKHSA. Sexually transmitted infections and screening for chlamydia in England: 2022 report 2023.++
- 4. GOV.UK. Gonorrhoea and syphilis at record levels in 2022 2023.++
- Shah D, Marfatia YS. Serological tests for syphilis. Indian Journal of Sexually Transmitted Diseases and AIDS 2019: volume 40, issue 2, pages 186-91.++ 10.4103/ijstd.IJSTD_86_19
- 6. Mitja O and others. Yaws. Lancet 2013: volume 381, issue 9868, pages 763-73.+ S0140-6736(12)62130-8 [pii];10.1016/S0140-6736(12)62130-8 [doi]
- Janier M and others. 2020 European guideline on the management of syphilis. Journal of the European Academy of Dermatology and Venereology 2020: volume 35, issue 3, pages 559-772.++ <u>https://doi.org/10.1111/jdv.16946open_in_new</u>
- 8. Heymans R and others. Clinical value of Treponema pallidum real-time PCR for diagnosis of syphilis. Journal of Clinical Microbiology 2010: volume 48, issue 2, pages 497-502.3+
- Larsen SA and others. Laboratory diagnosis and interpretation of tests for syphilis. Clinical Microbiology Reviews 1995: volume 8, issue 1, pages 1-21.+
- Pillay A. Centers for Disease Control and Prevention Syphilis Summit-Diagnostics and Laboratory Issues. Sex Transm Dis 2018: volume 45, issue 9S Suppl 1, pages S13-s6.++ 10.1097/olq.00000000000843
- Zhou J and others. An Updated Review of Recent Advances in Neurosyphilis. Frontiers in Medicine 2022: volume 9, pages 800383.++ 10.3389/fmed.2022.800383
- Marra CM and others. The rapid plasma reagin test cannot replace the venereal disease research laboratory test for neurosyphilis diagnosis. Sexually Transmitted Diseases 2012: volume 39, issue 6, pages 453-7.3+ 10.1097/OLQ.0b013e31824b1cde [doi];00007435-201206000-00010 [pii]

- Wohrl S, Geusau A. Neurosyphilis is unlikely in patients with late latent syphilis and a negative blood VDRL-test. Acta Dermato Venereologica 2006: volume 86, issue 4, pages 335-9.3- 10.2340/00015555-0092 [doi]
- 14. Marra CM and others. Cerebrospinal fluid abnormalities in patients with syphilis: association with clinical and laboratory features. The Journal of Infectious diseases 2004: volume 189, issue 3, pages 369-76.3- JID30874 [pii];10.1086/381227 [doi]
- Marks M and others. Diagnostic performance of PCR assays for the diagnosis of neurosyphilis: a systematic review. Sexually Transmitted Infections 2018: volume 94, issue 8, pages 585-8.1+ 10.1136/sextrans-2018-053666
- 16. GOV.UK. ISOSS syphilis report 2022 2022.++
- 17. Dorfman DH, Glaser JH. Congenital syphilis presenting in infants after the newborn period. The New England Journal of medicine 1990: volume 323, issue 19, pages 1299-302.**3-** 10.1056/NEJM199011083231902 [doi]
- Kelly M and others. The utility of the syphilis enzyme immunoassay IgM in the diagnosis of congenital syphilis. Diagnostic Microbiology and Infectious Disease 2020: volume 98, issue 3, pages 115152.1 10.1016/j.diagmicrobio.2020.115152
- 19. Cooper JM, Sánchez PJ. Congenital syphilis. Seminars in Perinatology 2018: volume 42, issue 3, pages 176-84.++ 10.1053/j.semperi.2018.02.005
- 20. Home Office. Anti-terrorism, Crime and Security Act. 2001. ++
- 21. Advisory Committee on Dangerous Pathogens. The Approved List of Biological Agents. Health and Safety Executive 2021. pages 1-39. ++
- 22. Health and Safety Executive. Blood-borne viruses in the workplace. Guidance for employers and employees. HSE. 2001. ++
- British Standards Institution (BSI). BS 5726:2005 Microbiological safety cabinets. Information to be supplied by the purchaser and to the vendor and to the installer, and siting and use of cabinets. Recommendations and guidance. 2005. pages 1-14. ++
- 24. British Standards Institution (BSI). BS EN12469 Biotechnology performance criteria for microbiological safety cabinets 2000. ++
- 25. Health and Safety Executive. Control of Substances Hazardous to Health Regulations. The Control of Substances Hazardous to Health Regulations 2002 (as amended). Approved Code of Practice and guidance L5 (sixth edition). HSE Books. 2013. ++

- 26. Official Journal of the European Communities. Directive 98/79/EC of the European Parliament and of the Council of 27 October 1998 on *in vitro* diagnostic medical devices 1998. pages 1-37. ++
- 27. World Health Organization. Guidance on regulations for the transport of infectious substances 2019-2020. WHO. 2019. ++
- 28. Centers for Disease Control and Prevention. Guidelines for Safe Work Practices in Human and Animal Medical Diagnostic Laboratories. MMWR Surveill Summ 2012: volume 61, pages 1-102.+
- 29. Department of Health. Health Protection Legislation (England) Guidance. pages 1-112. 2010. ++
- 30. The Welsh Assembly Government. Health Protection Legislation (Wales) Guidance. 2010. ++
- 31. Public Health England. Laboratory reporting to UKHSA: a guide for diagnostic laboratories. UKHSA 2022. pages 1-31. ++
- Health and Safety Executive, Advisory Committee on Dangerous Pathogens. Management and operation of microbiological containment laboratories. HSE. 2019. ++
- 33. Health and Safety Executive. Managing risks and risk assessment at work (accessed 28/07/2021). <u>https://www.hse.gov.uk/simple-health-safety/risk/index.htm</u>. ++
- 34. Scottish Government. Public Health (Scotland) Act. 2008. ++
- 35. Home Office. Public Health Act (Northern Ireland) 1967 Chapter 36. 1967. ++
- 36. The Royal College of Pathologists. The retention and storage of pathological records and specimens (5th edition). pages 1-59. 2015. ++
- 37. Health and Safety Executive. Risk assessment: A brief guide to controlling risks in the workplace. HSE. 2014. ++
- 38. Health Services Advisory Committee. Safe working and the prevention of infection in clinical laboratories and similar facilities. Books. H 2003. ++
- 39. Department for Transport and others. Transport of infectious substances UN2814, UN2900 and UN3373 Guidance note number 17/2012 (revision 7).
 2013. ++